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## HOW EXPERIMENTAL PHYSIOLOGY AND ECOLOGICAL NICHE MODELLING CAN INFORM THE MANAGEMENT OF MARINE BIOINVASIONS?

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## ABSTRACT

Marine bioinvasions are increasing worldwide by a number of factors related to the anthroposphere, such as higher ship traffic, climate change and biotic communities' alterations. Generating information about species with high invasive potential is necessary to inform management decisions aiming to prevent their arrival and spread. *Grateloupia turuturu*, one of the most harmful invasive macroalgae, is capable of damaging ecosystem functions and services, and causing biodiversity loss. Here we developed an ecological niche model using occurrence and environmental data to infer the potential global distribution of *G. turuturu*. In addition, ecophysiological experiments were performed with *G. turuturu* populations from different climatic regions to test predictions regarding invasion risk. Our model results show high suitability in temperate and warm temperate regions around the world, with special highlight to some areas where this species still doesn't occur. Thalli representing a potential temperate region origin, were held at 10, 13, 16, 20 and 24°C, and measurements of optimal quantum field ( $F_v/F_m$ ) demonstrated a decrease of photosynthetic yield in the higher temperature. Thalli from the population already established in warm temperate South Atlantic were held at 18, 24 and 30°C with high and low nutrient conditions. This material exposed to the higher temperature demonstrated a drop in photosynthetic yield and significant reduction of growth rate. The congregation of modelling and physiological approach corroborate the invasive potential of *G. turuturu* and indicate higher invasion risk in temperate zones. Further discussions regarding management initiatives must be fostered to mitigate anthropogenic transport and eventually promote eradication initiatives in source areas, with special focus in the South America. We propose that this combined approach can be used to assess the potential distribution and establishment of other marine invasive species.

Keywords: *Grateloupia turuturu*, invasive marine algae, populational ecology, ecophysiology, anthropogenic impacts.

## 1. INTRODUCTION

Marine bioinvasions are receiving great attention as many of them are producing negative socio-environmental-economic impacts worldwide (Katsanevakis et al., 2014). Offshore oil exploitation, aquaculture activities and ballast water are major vectors responsible for growing events of non-indigenous species arrival and spread (Creed et al., 2017; Hulme, 2009). Climate change further facilitates the dispersal of marine organism by increasing the frequency and intensity of extreme climatic/oceanographic events (Sissini et al., 2017, 2014; Ummenhofer and Meehl, 2017). A diverse and abundant array of vectors for the transport of organisms across biogeographic boundaries coincides with disrupted native communities of marine organisms, caused by both global (ocean acidification and warming) and local stressors (coastal pollution, overfishing) (Shea, 2002), which is facilitating the establishment of non-indigenous species on coastlines all around the world (Occhipinti-Ambrogi 2007). Williams and Smith (2007) estimated that there are 277 introduced seaweed species around the globe, with the Mediterranean and the North-East Atlantic being the most highly invaded regions. The geographical distribution of marine macroalgae is determined by their physiological tolerance to environmental conditions (Breeman, 1988; Hoek, 1982), enabling the prediction of suitable habitats for invasive species (Elith et al., 2006; Marcelino and Verbruggen, 2015). Ecological Niche Models (ENM) relate occurrence data with environmental gradients to produce habitat suitability maps (Peterson, 2003). An ability to assess the risk of invasion and the threats posed by foreign organisms helps managers and stakeholders put preventive measures in place and optimize biosecurity monitoring (Marcelino

and Verbruggen, 2015). Eradication of established populations is costly and usually ineffective (Bax et al., 2001; D’Antonio and Meyerson, 2002). The known occurrence of species informs predictive distribution models, which can be misleading since this depends on sampling effort and identification expertise which varies widely depending on the species and regions investigated (Verbruggen et al., 2009). ENM do not consider the physiological traits of species when modelling distributions, likely missing valuable information about the mechanisms and processes allowing the presence and establishment of populations (Marcelino and Verbruggen, 2015). Therefore, physiological information can be used to validate and further improve ENMs to create a more accurate prediction of habitat suitability (Martínez et al., 2015).

Considering species traits related with dispersal, establishment and ecological impact, the fifth most dangerous invasive alga in a global perspective is *Grateloupia turuturu* Yamada (1941) (Nyberg and Wallentinus, 2005), commonly known as the “Devil’s tongue weed” (Mulas and Bertocci, 2016). *G. turuturu* can displace native species in rocky intertidal habitats (Freitas et al., 2016; Mulas and Bertocci, 2016), has advantage establishing in recently perturbed environments, where native assemblages are disrupted (Mulas and Bertocci, 2016), and has been reported for reducing the abundance and richness of invertebrates (Janiak and Whitlatch, 2012; Jones and Thornber, 2010). Native to Japan, *G. turuturu* has been documented as an invasive species in the United States (Balcom and Yarish, 1994; Mathieson et al., 2008), Mexico (Aguilar-Rosas et al., 2012), New Zealand (D’Archino et al., 2007), France (Simon et al., 2001), Italy (Cecere et al., 2011), Portugal (Araújo et al., 2011), Australia (Saunders and Withall, 2006), and Israel (Tsiamis et al., 2015). This species was recently registered for the coast of South Brazil, being the first record of this species in the South Atlantic (Azevedo et al., 2015). The transport of oyster for aquaculture and ballast water are the main probable vectors of *G. turuturu* introduction in the

Atlantic and Mediterranean (Araújo et al., 2011; Harlin and Villalard-Bohnsack, 2001). Important dispersion vectors may also be yacht and ship hulls, as specimens have been found attached to such boats in France (Simon et al., 2001). *G. turuturu* has been described to tolerate a wide range of temperature (from 4 to 28°C), salinity (from 15 to 37), nutrients (Simon et al., 2001) and can be found down to a depth of 7m (Sweet 2011). Liu and Pang (2010) described lower photosynthetic activity in *G. turuturu* when exposed to freezing, which further explains why this species is mostly found, until now, in temperate regions. However, even with the description of *G. turuturu* being tolerant to a great spectrum of environmental conditions in the North Atlantic and Pacific, there is no descriptions of the South Atlantic population tolerance to the local oceanographic conditions, nor experiments that describe the biochemical processes of its ecophysiological performance. For this reason, ecophysiological experiments are necessary to better understand the influence of temperature and nutrients in *G. turuturu* physiology and assess its probability of success in regions with conditions similar to the native range.

In this study we developed an ENM to estimate the global potential distribution of *G. turuturu*. In addition, we determined the ecophysiological thresholds of *G. turuturu* as a tool to infer about the species resilience and temperature tolerance. Our hypothesis is that *G. turuturu* will present higher habitat suitability and physiological performance in temperate regions. By combining ENMs and the physiological thresholds evaluation, we desire to propose this associated approach to foster coastal management strategies related not only to *G. turuturu* bioinvasion, but also to invasions by other marine species.

## 2. MATERIALS AND METHODS

### 2.1 Ecological Niche Modelling

To develop the ENM for *G. turuturu*, six environmental benthic data layers (i.e., along the bottom of the ocean) were extracted from Bio-ORACLE (Assis et al., 2017b; Tyberghein et al., 2012) based on the biological relevance of macroalga (Assis et al., 2017a, 2014; Neiva et al., 2015) (environmental predictors used in the model are described in Table 1). Depth was not included as a predictor as it has no direct biological meaningfulness in the ecophysiological response of macroalgae, and many previous works modelling marine macroalgae also do not include it (Jueterbock et al., 2016, 2013; Tyberghein et al., 2012). Bio-ORACLE data has been widely used for marine ecological modelling (Marcelino and Verbruggen, 2015) since it contains climatic and geophysical data with a global spatial resolution of 0.08° (9.2 km). Detailed georeferenced occurrence data from both invaded and native ranges was obtained from the public Global Biodiversity Information Facility (GBIF, 2018), and literature data (available in the supplementary material ESM1) (e.g. Tyberghein et al. 2012; Escobar et al. 2016). The inclusion of the invaded range in the model increases model accuracy, as models trained using data only from the native range usually fail to predict the full invasion extent of a species (Beaumont et al., 2009; Broennimann and Guisan, 2008). Duplicated occurrences, outside marine range or too close to each other (5 km<sup>2</sup>) were excluded to prevent biases and reduce spatial autocorrelation in the analyses, remaining 106 records (Figure 1). The radius of 5 km<sup>2</sup> was chosen as a conservative distance of about half of the resolution of the climatic data used in the model, seeking spatial independence between the environmental data of occurrences (Phillips et al. 2009; Kramer-Schadt et al. 2013). The modelling approach combined multiple cross-validation interactions using independent datasets with Boosted Regression Trees (BRT), a statistical algorithm able to fit complex non-linear relationships between a response (occurrence records) and predictor variables (environmental data), while avoiding overfitting through optimized parameterization procedures

and specific forcing of monotonic responses (Elith, Leathwick, & Hastie, 2008). BRT algorithm is known for high predictive performance (De'ath, 2007; Neiva et al., 2014), improving its performance by combining many models for prediction, which makes it a more accurate algorithm when compared to other regression methods (Elith, Leathwick, & Hastie, 2008). In the lack of absence data, presence-only modelling requires the generation of pseudo-absences or background information (e.g., Maxent; Phillips et al. 2006). This information was produced using the method developed by Senay et al. (2013) for improved ecological modelling, which comprise the calculation of environmentally dissimilar regions to the presence points and the random selection of pseudo-absences in clustered climatic predictors to reduce the use of redundant information. To account for likely sources of sampling bias, pseudo-absences were chosen from a kernel probability surface developed with the records of occurrence over a grid with the same resolution of environmental data (e.g., Phillips et al. 2009; Assis et al. 2018a)

Cross-validation interactions were developed by partitioning species records (i.e., presences and also pseudo-absences) into 10 independent longitudinal bands to evaluate the performance of models (Wenger and Olden, 2012). Models fitted with one withheld band at a time, and True skill statistics (TSS) compared this data to the outcomes of a predicted surface of distribution reclassified to reflect presence and absences (binomial) with a threshold maximizing TSS. Over-fitting in the models was reduced by tuning the algorithm parameters (i.e., learning rate, number of trees and tree complexity; see Elith et al. 2008; Assis et al. 2016, 2018a for details) and also by forcing monotonic responses (positive for minimum nutrients, salinity, light and temperature; negative for maximum temperatures). Final predictions using optimal parameters were reclassified to binomial responses and its final accuracy was reported with AUC (Area Under the receiver operating characteristic Curve, Fielding and Bell 1997), sensitivity (true positive rate)



and TSS (True Skill Statistics, Allouche et al. 2006). The significance of models was determined by inferring the relative contribution of each environmental predictor to the performance of the models (Elith et al., 2008). Physiological limits points (maximum and minimum, depending on the predictor) were determined by extracting the individual fitting function of each predictor alone, while fixing the alternative predictors in their average (Elith et al., 2008; Marcelino and Verbruggen 2015).

ENM analyses were performed with R (R Development Core Team, 2017) using the packages dismo, gbm, parallel, raster, SDMTools, and sm.

## 2.2 Ecophysiological experiments

Two populations of *G. turuturu* were ecophysiotogically evaluated considering different acclimatization histories. A warm temperate adapted population (WAP) was sampled (approximately 100 g of fresh weight (FW)) on August 22<sup>nd</sup>, 2016 from the intertidal zone of Itapirubá beach (28°20'14.32"S, 48°42'4.87"W), in the city of Imbituba, Santa Catarina, Southern Brazil. A temperate adapted population (TAP), approximately 60 g FW, was also sampled in Saint Malo beach (48°39'32.8"N 2°00'24.6"W), France, on February 20<sup>th</sup>, 2017, region representing a possible source of *G. turuturu* germlings through oyster seeds or ballast water. Since a determined biomass of *G. turuturu* was collected, which included individual blades and blades attached together by a holdfast, all samples were separated in individual blades. The sampled blades of both populations presented around 6 cm height and 2 cm in width, were not fertile, collected in winter, immediately transported in plastic containers with seawater to the lab, and manually cleaned from epiphytes or associated fauna after sampling.

To determine the control conditions for each of the experimental designs, we considered the different adaptation conditions of each population. Therefore, the temperate and warm temperate populations were exposed to conditions observed in their respective occurrence regions (~10°C in France in February (Dubois et al., 2007), and ~18°C in Imbituba in August (Gouvêa et al., 2017)). Both populations were cultivated in 250 mL Erlenmeyer flasks, in UV sterilized seawater enriched with Von Stosch medium in a 50% concentration, without EDTA chelating salt (adapted from Edwards 1972) until the beginning of the experiment. To prevent the lack of essential nutrients water was replaced daily. Aeration was provided individually in the flasks, also stirring the water in each flask to disrupt the diffusive boundary layer. The temperature was adjusted by heaters (Radical 1 HP, Brazil) positioned in trays with water, where the treatment flasks were kept in a water bath. Each tray contained one data logger (HOBO Pendant Temperature/Light Data Loggers, Onset, Bourne, USA) that recorded the water temperature every 10 minutes. A 12-hour light/dark photoperiod was used, and irradiance was provided by fluorescent lamps with an average of  $65 \pm 5 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ .

The experimental design performed in Brazil had as main purpose to observe short-term ecophysiological effects of temperature and nutrients, key environmental drivers selected according to conditions found in adjacent tropical environments where the species has not been reported yet (Gouvêa et al., 2017). The WAP sampled blades were brought to the laboratory, and acclimated for 21 days to laboratory irradiance, maintaining field temperature (18°C). Initial and final 1g FW samples were frozen with liquid nitrogen for the analysis of biochemical descriptors. The experiment lasted for seven days. The experimental setup in Brazil utilized three temperatures (18, 24 and 30°C) and two nutrient conditions: one without medium culture (Low Nutrient Concentration – LNC) and another with 50% of Von Stosch (VS) medium (High Nutrient

Concentration – HNC), resulting in 6 factorial treatments. Each treatment contained 10 replicas ( $n = 10$ ), consisting of individual blades of *G. turuturu* weighted to sum 1g FW per Erlenmeyer flask.

The experiment performed with the temperate adapted population, a potential source of *G. turuturu*, has as main purpose the investigation of the short-term ecophysiological effects of temperature, an environmental driver traditionally related with the distribution constraints of marine algae (Hoek, 1982). The chosen temperature values used in the experiment are present in the region of the North Atlantic where the algae were sampled (Dubois et al., 2007), except for the higher temperature used. The higher temperature used was chosen to overlap with conditions observed in the warm temperature region, providing the necessary scenario for the evaluation of the potential ecophysiological continuum, contribution to the discussion about invasive potential of *G. turuturu*. For the TAP experiment realized in France, *G. turuturu* blades were acclimated for three days at 10°C, temperature observed in the water of the sampling location. *G. turuturu* was exposed for 60 hours to 5 different temperatures: 10 (control), 13, 16, 20 and 24°C. Each treatment contained 6 replicas ( $n = 6$ ), also consisting of individual blades of *G. turuturu* weighted to sum 1g FW. The experiment duration was limited by laboratory structure and the availability at Saint Malo. Figure S1 on the supplementary material ESM2 illustrates the different experimental designs of both populations.

### 2.3 Physiological analyses

Physiological analysis performed in both populations were measurements of optimal quantum field (Fv/Fm). Growth rate and photosynthetic pigments were only evaluated in the WAP experiment, as the biomass sent to Brazil lost minimum preservation conditions due to

transportation delay. Considering the relevance of Fv/Fm as a descriptor, we chose to keep the results obtained in the TAP experiment.

### 2.3.1 Photosynthetic yield

In the WAP experiment, effective quantum yield (Fv/Fm) was estimated by the fluorescence of *in vivo* chlorophyll *a*. Three measurements of Fv/Fm were made in each replica in different parts of the thallus, resulting in 3 subreplicas of each one of the 10 replicas. Measurements of Fv/Fm were taken with a pulse-amplitude modulated chlorophyll *a* fluorometer (Diving PAM – Walz Germany) after acclimation in the dark for 20 minutes in the first day, then every two days, and in the last day of the experiment (for further explanation of the methodology followed, see: de Ramos et al., 2018; Figueroa et al., 2006). For the TAP experiment, measurements of Fv/Fm were made in the dark only in the end of the 60 hours.

### 2.3.2 Pigment extraction

At the end of the experiment, all the replicates from the WAP experiment were frozen and used for pigments quantification. Each replica was divided in 3 portions of 100 mg, allowing the reading of 3 subreplicas, which resulted in 30 extractions for each treatment. The extraction followed Schubert et al. (2011), where phycobiliproteins, chlorophyll *a* and total carotenoids were extracted from the same 100 mg biomass sample grinded in liquid nitrogen. Phycobiliproteins were extracted first with phosphate buffer (pH 6.8) and kept in the dark at 4°C for 2 hours until centrifugation for 4 minutes at 13000 rpm in the MiniSpin centrifuge (Eppendorf) at room temperature. Subsequently, the supernatant was used to determine the concentration of phycobiliproteins according to Beer and Eshel (1985). Afterwards, methanol 100% was placed on

the same biomass and kept in the dark at 4°C overnight, and then centrifuged as stated before. Then, chlorophyll and total carotenoids were quantified according to Torres et al. (2014). Absorbance was determined in the SpectraMax Paradigm Multi-Mode Microplate Reader (Molecular Devices), of the Laboratório Multiusuário de Estudos em Biologia (LAMEB).

### 2.3.3 Growth rate (GR)

To determine the daily growth rate (GR), replicates were spun for 15 seconds in a salad spinner to reduce the excess of water, and then weighted in an analytical balance. The replicates were weighted in the first and last days to determine the GR by the following equation (Lignell and Pedersén, 1989):

$$GR (\% \cdot \text{day}^{-1}) = \left[ \left( \frac{W_t}{W_i} \right)^{1/t} \right] \times 100\% \quad (1)$$

Where  $W_i$  and  $W_t$  are respectively the initial and final fresh weight after  $t$  days ( $t = 7$  days).

### 2.4 Statistical analysis

For the WAP experiment, the independent variables considered were time (beginning and end), nutrient concentration (HNC and LNC) and temperature (18, 24 and 30°C). For Fv/Fm, time was considered for the 1st, 3rd, 5th and 7th day. For significant differences, post-hoc test of Tukey was used to identify significantly different groups ( $p < 0.05$ ). The TAP experiment had as variables only the different temperatures, 10, 13, 16, 20 and 24°C, with 6 replicates ( $n=6$ ).

Each treatment of the WAP experiment had 10 replicates ( $n=10$ ), which were used for the analysis of GR. However, in the reading of pigments and Fv/Fm, some of the replicas presented negative values or values too close to zero, representing errors in the readings. This required the removal of 3 replicas in each treatment (resulting in a  $n=7$ ), to maintain the same number of

replicates for statistical analysis. Before analysis, data were tested for normality and residuals for half-normality. Statsoft Statistica 13.0 was used to perform multifactorial Analysis of Variance (ANOVA,  $p < 0.05$ ). Due to the removal of outliers, the data did not require transformation, as it attained the normality requirements for the ANOVA parametric tests calculated.

### 3. RESULTS

#### 3.1 Ecological Niche Modelling

The final predictive model had high performance (AUC: 0.96; Sensitivity: 0.95; TSS: 0.93) and showed that *G. turuturu* has great habitat suitability in temperate regions of Northern Pacific, Northern Atlantic, South America, Australasia (Oceania) and a small portion of Southern Africa (Fig. 2). Light at bottom and maximum ocean temperature were the most important predictors to the ecological niche of *G. turuturu*, while nutrients (as nitrates and phosphates) were the least important (Table 1). Response curves showed habitat suitability with light above  $4.4 \text{ E} \cdot \text{m}^{-2} \cdot \text{yr}^{-1}$  and ocean temperature below  $26.55^\circ\text{C}$ , however, with a starting drop at  $21.23^\circ\text{C}$  (Fig. 3). Response curves of the other variables used in the model are available in the supplementary material ESM2 (Figures S2, S3, S4, S5 and S6).

#### 3.2 Ecophysiological experiments and physiological analyses

##### 3.2.1 Photosynthetic yield

In the WAP experiment, there was significant interaction ( $p < 0.05$ ) between temperature, time and nutrient concentration for Fv/Fm (Table 2). A decline in Fv/Fm was noticeable as the temperature increases and through the experiment time in WAP (Fig. 4). The treatment of  $30^\circ\text{C}$  with HCN showed the lowest values in the fifth and seventh day of treatment in comparison with

the treatments with lower temperatures. For 30°C with LNC, there was a smaller decline in Fv/Fm, but still presented Fv/Fm values lower than the treatments of 24 and 18°C. Similarly, there was a contrast between the LNC and HNC treatments at 24°C, with lower values of Fv/Fm observed at HNC. In France, statistical analysis of TAP experiment results of Fv/Fm showed significant difference between temperatures (Table 2). The response of *G. turuturu* to the short-term exposure in the TAP experiment to different temperatures was of lower Fv/Fm in the treatment of 24°C (Fig. 5).

### 3.2.2 Photosynthetic pigments

No significant interaction ( $p>0.05$ ) was found for both GR and pigments with time, temperature, and nutrient concentration (i.e., no three-factorial interaction for GR and pigments). However, regarding chlorophyll *a*, total carotenoids and GR, when the effects of nutrient concentration are taken out, the interaction between time and temperature was significant ( $p<0.05$ ). For phycoerythrin and chlorophyll *a* the interaction between the effects of time and nutrient concentrations was significant (Table 3). For chlorophyll, the treatments of 24°C and 30°C in the end of the experiment were statistically distinct from the samples of the beginning and 18°C (Fig. 6a). The higher concentration of chlorophyll was found in 30°C, followed by 24°C. Similarly, the higher concentration of total carotenoids was found in the last day at 30°C, and all the other treatments had the same lower concentration (Fig. 6b). Additionally, for chlorophyll there was statistical difference between different nutrient concentrations. On the end of the experiment, the HNC treatment presented higher values of chlorophyll, as in LNC the concentration remained close to the one in the beginning of the experiment (Fig. S7 on the supplementary material ESM2). On the other hand, there was a decrease in phycoerythrin concentration in the last day in LNC,

while at the treatment of HNC remained constant throughout the experiment (Fig. 7).  
Concentration of phycocyanin was not detectable.

### 3.2.3 Growth rate

For daily growth rate, only temperature and time were significant (Table 3). The algae  
exposed to 30°C showed lower GR when compared to the algae in lower temperatures (Figure 6c).

## 2.4 DISCUSSION

The investigation of the potential geographic distribution through ENM and physiological  
thresholds tested through experiments of *G. turuturu* indicated higher habitat suitability of this  
invasive species in temperate and warm-temperate regions around the world. Experimental results  
demonstrated a decrease in photosynthetic yield in algae exposed to higher temperatures, along  
with a stress response observed in pigments and growth rate. At the same time, ENM showed a  
possible expansion of the distribution of *G. turuturu* to new countries around the globe, especially  
in regions with temperatures close to the ones that the algae better adapted in the experiments.

The variables that most contributed to explain the distribution of *G. turuturu* were the  
availability of light at bottom and maximum ocean temperatures. Light at bottom is determined  
with an exponential decay function of photosynthetic active radiation, diffuse attenuation  
coefficient and depth (Assis et al., 2017b) and allows restricting the species vertical distribution.  
Indeed, the  $4.04 \text{ E. m}^{-2} \cdot \text{yr}^{-1}$  threshold is comparable with the minimum annual irradiance inferred  
for other species of algae (Gattuso et al., 2006). The limitation of 26.55°C maximum temperature  
inferred in ENM was also in line with previous descriptions of Simon et al. (2001) for the species  
occurrence in Brittany (France), where waters can reach up to 28°C in summer. The relative



discrepancy in these values can arise from the short term variations of temperature in the water pools where Simon et al. (2001) described the occurrences, which are not captured in the averaged climatic layers used for modelling. In fact, our experiment results show, the warm temperate population of *G. turuturu* can maintain its photosynthetic yield for at least three days in this temperature, demonstrating that this alga can survive to short-time exposure to high temperatures.

In addition, the temperate population experiment demonstrated *G. turuturu* similar Fv/Fm responses to temperature as the response curve from the model, with higher photosynthetic yield in 10, 13, 16 and 20°C. The experiment with the blades collected in Brazil also showed higher Fv/Fm in the lower temperature used. These results indicate that both populations of *G. turuturu* present better physiological performance in regions with mild temperatures, i.e. temperate climates. Interestingly, in higher temperatures the two populations responded differently. As stated above, the WAP population presented tolerance to a short-time exposure to 30°C, and to 24°C for the duration of the experiment (LNC only), while the TAP population had a significant drop in performance in 24°C. Although the different responses may be relative to the fact that these are two distinct invasive populations (Allendorf and Lundquist, 2003), both demonstrated the tendency to experience a drop in Fv/Fm in higher temperatures. The higher temperature used in the warm temperate experiment demonstrated that *G. turuturu* will also present reduced growth rate in regions with average temperatures around 30°C. Even with the higher chlorophyll content, *G. turuturu* was not able to maintain its photosynthetic yield in 30°C. Furthermore, the higher concentration of total carotenoids showed a stress response of the algae to the higher temperature but did not prevent the reduction of growth. However, previous studies of the species demonstrated that younger stages of *G. turuturu*'s life cycle have a wider temperature tolerance range than older stages, with germinating growth being the same at 15°C and 30°C (Kraemer et al., 2017). For this

reason, juvenile stages and/or spores may be more aggressive in a bioinvasion process if transported by an oceanographic vector. And once in a new habitat, the acclimation conditions during the invasive process may be determinant in the success of establishment, when temperature limits are between the ones here evaluated for adult blades.

The ecophysiological experiment in Brazil showed that *G. turuturu* can tolerate a wider range of nutrient concentration without reducing its photosynthetic yield in 18°C, which was expected based on previous descriptions of this algae (Simon et al., 2001). The region where the species is found in southern Brazil is characterized by the presence of upwelling of colder, nutrient-rich waters from the South Atlantic Central Water, and during winter receives influence from the nutrient-rich La Plata river plume and the cold waters of the Sub-Antarctic currents (Pereira et al., 2009). Simultaneously, Martins et al. (2012) reported the algae diversity decrease in southern Brazil caused by urbanization and coastal pollution, damaging the resilience of the benthic community. Consequently, the local conditions of this region meet the suitable conditions demonstrated by the experiment and the model for the establishment and expansion of *G. turuturu* occurrence. Hence, countries with similar coastal characteristics with no register of the species must be aware and act to prevent the arrival and establishment of *G. turuturu*.

Our results allow the affirmation that non-fertile adult blades of *G. turuturu* will likely not have success in expanding its niche to regions with an average temperature around 30°C or survive to heatwaves (short periods with an average of 5 days of anomalous warming of the ocean) (Hobday et al., 2016) in regions with milder temperatures. As for temperate and warm-temperate regions, our model predicts habitat suitability for *G. turuturu* for temperatures reaching up to approx. 26°C, and the WAP experiment results demonstrated that this alga can maintain its photosynthetic yield and growth at 24 °C. However, if in 24°C an event of introduction of a higher

amount of nutrients in the water column occurs, *G. turuturu* may suffer a drop in photosynthetic yield depending on the exposure period. Consequently, the invasive potential of this alien species in regions with an average temperature around 24°C may be smaller than its potential in colder water regions, as noted in the 18°C treatment, where *G. turuturu* had no response to the higher nutrient concentration.

Temperature was chosen as one of the variables in the experiments and model due to its importance in determining the geographic distribution of macroalgae (Hoek, 1982). The changes in the average sea surface temperature caused by climate change are characterized as a global stressor to autochthonous communities. *G. turuturu*, as demonstrated in this study, can have advantage in such scenario, expanding its distribution to locations with predicted regional cooling (e.g. northern Chile and South Africa, Falvey and Garreaud 2009; Bolton and Anderson 2012) or warming of cold-water regions (Poloczanska et al., 2013; Vergés et al., 2014). Models as the one here used are the best approach to translating future climatic changes into ecological responses, guiding strategies of conservation and management (Wiens et al., 2009). Moreover, the effects of a local stressor, higher nutrient concentration, were assessed in *G. turuturu* through ecophysiology evaluation. By combining both approaches, we could determine the possible geographic expansion of *G. turuturu* under both local and global stressors.

Habitat models for invasive seaweeds need a high transferability, expanding predictions to regions where the species could potentially invade. Verbruggen et al. (2013) demonstrated that for global predictions the most important procedure to increase model transferability is choosing fewer predictors with high ecological significance for the species. Furthermore, the use of ecophysiological knowledge about the species is highly advisable to assess the significance of

411 predictions (Araújo and Guisan, 2006). Accordingly, we used ENM and fewer variables to model  
412 the global distribution of *G. turuturu* together with the evaluation of its ecophysiology.

413         With increasing environmental changes and availability of transport vectors, it is essential  
414 the use and improvement of approaches capable of predicting possible new events of bioinvasion,  
415 as done in this work. Invasive species represent great harm to ecosystem services, with provision  
416 services being the most affected by them, such as food provision and genetic variability (Vilà et  
417 al., 2010). It becomes clear that these predicting tools and the knowledge of invasive species and  
418 their habitats must be used as the foundation of international regulatory agreements, pursuing the  
419 protection of natural resources and biodiversity (Hulme et al., 2008). The results found in this  
420 study enable stakeholders and policymakers to discuss and foster management actions regarding  
421 prevention or eradication in case of *G. turuturu* expanding its distribution to new regions,  
422 especially for South America where there are few registers of its occurrence (Azevedo et al., 2015).  
423 Therefore, the present work takes place as an important tool of bioinvasions management in a  
424 scenario of quick changes in environmental conditions and dispersion patterns, in which the delay  
425 of response by international and local governments can cost the planet ecosystem functions and  
426 services that it relies upon.

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## 2.5 REFERENCES

- Aguilar-Rosas, L.E., Boo, S.M., Kim, K.M., Aguilar-Rosas, C. V, 2012. First record of the Japanese species *Grateloupia turuturu* (Halymeniaceae, Rhodophyta) in the Mexican Pacific coast. *Hidrobiologica* 22, 189–194.
- Allendorf, F.W., Lundquist, L.L., 2003. Introduction: Population biology, evolution, and control of Invasive Species. *Conserv. Biol.* 17, 24–30. <https://doi.org/10.1046/j.1523-1739.2003.02365.x>
- Allouche, O., Tsoar, A., Kadmon, R., 2006. Assessing the accuracy of species distribution models: Prevalence, kappa and the true skill statistic (TSS). *J. Appl. Ecol.* 43, 1223–1232. <https://doi.org/10.1111/j.1365-2664.2006.01214.x>
- Araújo, M.B., Guisan, A., 2006. Five (or so) challenges for species distribution modelling. *J.*

- 456 Biogeogr. 33, 1677–1688. <https://doi.org/10.1111/j.1365-2699.2006.01584.x>
- 457 Araújo, R., Violante, J., Pereira, R., Abreu, H., Arenas, F., Sousa-Pinto, I., 2011. Distribution  
458 and population dynamics of the introduced seaweed *Grateloupia turuturu* (Halymeniaceae,  
459 Rhodophyta) along the Portuguese coast. *Phycologia* 50, 392–402.  
460 <https://doi.org/10.2216/10-65.1>
- 461 Assis, J., Araújo, M.B., Serrão, E.A., 2017a. Projected climate changes threaten ancient refugia  
462 of kelp forests in the North Atlantic. *Glob. Chang. Biol.* 24, e55–e66.  
463 <https://doi.org/10.1111/gcb.13818>
- 464 Assis, J., Coelho, N.C., Lamy, T., Valero, M., Alberto, F., Serrão, E.Á., 2016. Deep reefs are  
465 climatic refugia for genetic diversity of marine forests. *J. Biogeogr.* 43, 833–844.  
466 <https://doi.org/10.1111/jbi.12677>
- 467 Assis, J., Serrão, E.A., Claro, B., Perrin, C., Pearson, G.A., 2014. Climate-driven range shifts  
468 explain the distribution of extant gene pools and predict future loss of unique lineages in a  
469 marine brown alga. *Mol. Ecol.* 23, 2797–2810. <https://doi.org/10.1111/mec.12772>
- 470 Assis, J., Tyberghein, L., Bosch, S., Verbruggen, H., Serrão, E.A., De Clerck, O., 2017b. Bio-  
471 ORACLE v2.0: Extending marine data layers for bioclimatic modelling. *Glob. Ecol.*  
472 *Biogeogr.* 27, 277–284. <https://doi.org/10.1111/geb.12693>
- 473 Azevedo, C.A.A. de, Cassano, V., Horta, P.A., Batista, M.B., de Oliveira, M.C., 2015. Detecting  
474 the non-native *Grateloupia turuturu* (halymeniales, rhodophyta) in southern Brazil.  
475 *Phycologia* 54, 1–5. <https://doi.org/10/2216.15-25.1>
- 476 Balcom, N., Yarish, C., 1994. *Grateloupia turuturu*: A Red Seaweed Invading Long Island

- 477 Sound.
- 478 Bax, N., Carlton, J.T., Mathews-Amos, A., Haedrich, R.L., Howarth, F.G., Purcell, J.E., Rieser,  
 479 A., Gray, A., 2001. The control of biological invasions in the world's oceans. *Conserv.*  
 480 *Biol.* 15, 1234–1246. <https://doi.org/10.1046/j.1523-1739.2001.99487.x>
- 481 Beaumont, L.J., Gallagher, R. V., Thuiller, W., Downey, P.O., Leishman, M.R., Hughes, L.,  
 482 2009. Different climatic envelopes among invasive populations may lead to  
 483 underestimations of current and future biological invasions. *Divers. Distrib.* 15, 409–420.  
 484 <https://doi.org/10.1111/j.1472-4642.2008.00547.x>
- 485 Beer, S., Eshel, A., 1985. Determining phycoerythrin and phycocyanin concentrations in aqueous  
 486 crude extracts of red algae. *Mar. Freshw. Res.* 36, 785–792.  
 487 <https://doi.org/10.1071/MF9850785>
- 488 Bolton, J., Anderson, R., 2012. South African kelp moving eastwards: the discovery of *Ecklonia*  
 489 *maxima* (Osbeck) Papenfuss at De Hoop Nature Reserve on the south coast of South Africa.  
 490 *African J. Mar. Sci.* 34, 147–151. <https://doi.org/10.2989/1814232X.2012.675125>
- 491 Breeman, A.M., 1988. Relative importance of temperature and other factors in determining  
 492 geographic boundaries of seaweeds: Experimental and phenological evidence. *Helgoländer*  
 493 *Meeresuntersuchungen* 42, 199–241. <https://doi.org/10.1007/BF02366043>
- 494 Broennimann, O., Guisan, A., 2008. Predicting current and future biological invasions: Both  
 495 native and invaded ranges matter. *Biol. Lett.* 4, 585–589.  
 496 <https://doi.org/10.1098/rsbl.2008.0254>
- 497 Cecere, E., Moro, I., Wolf, M.A., Petrocelli, A., Verlaque, M., Sfriso, A., 2011. The introduced

- 498 seaweed *Grateloupia turuturu* (Rhodophyta, Halymeniales) in two Mediterranean  
 499 transitional water systems. Bot. Mar. 54, 23–33. <https://doi.org/10.1515/BOT.2011.009>
- 500 Chow, F., Furlan, C.M., Torres, P.B., Mercadante, A., Santos, D.Y.A.C. dos, Mandelli, F., 2014.  
 501 Standardization of a protocol to extract and analyze chlorophyll a and carotenoids in  
 502 *Gracilaria tenuistipitata* Var. Liui. Zhang and Xia (Rhodophyta). Brazilian J. Oceanogr. 62,  
 503 57–63. <https://doi.org/10.1590/s1679-87592014068106201>
- 504 Creed, J.C., Fenner, D., Sammarco, P., Cairns, S., Capel, K., Junqueira, A.O.R., Cruz, I.,  
 505 Miranda, R.J., Carlos-Junior, L., Mantelatto, M.C., Oigman-Pszczol, S., 2017. The invasion  
 506 of the azooxanthellate coral *Tubastraea* (Scleractinia: Dendrophylliidae) throughout the  
 507 world: history, pathways and vectors. Biol. Invasions 19, 283–305.  
 508 <https://doi.org/10.1007/s10530-016-1279-y>
- 509 D’Antonio, C., Meyerson, L.A., 2002. Exotic Plant Species as Problems and Solutions in  
 510 Ecological Restoration: A Synthesis. Restor. Ecol. 10, 703–713.  
 511 <https://doi.org/10.1046/j.1526-100X.2002.01051.x>
- 512 D’Archino, R., Nelson, W.A., Zuccarello, G.C., 2007. Invasive marine red alga introduced to  
 513 New Zealand waters: first record of *Grateloupia turuturu* (Halymeniaceae, Rhodophyta).  
 514 New Zeal. J. Mar. Freshw. Res. 41, 35–42. <https://doi.org/10.1080/00288330709509894>
- 515 De’ath, G., 2007. Boosted Trees for Ecological Modeling and Prediction. Ecology 88, 243–251.
- 516 de Ramos, B., da Costa, G.B., Ramlov, F., Maraschin, M., Horta, P.A., Figueroa, F.L., Korbee,  
 517 N., Bonomi-Barufi, J., 2018. Ecophysiological implications of UV radiation in the  
 518 interspecific interaction of *Pyropia acanthophora* and *Grateloupia turuturu* (Rhodophyta).



- 519 Mar. Environ. Res. <https://doi.org/10.1016/j.marenvres.2018.11.014>
- 520 Dubois, S., Comtet, T., Retière, C., Thiébaud, E., 2007. Distribution and retention of *Sabellaria*  
 521 *alveolata* larvae (Polychaeta: Sabellariidae) in the Bay of Mont-Saint-Michel, France. Mar.  
 522 Ecol. Prog. Ser. 346, 243–254. <https://doi.org/10.3354/meps07011>
- 523 Edwards, P., 1972. Cultured red alga to measure pollution. Mar. Pollut. Bull. 3(12), 184–188.  
 524 [https://doi.org/10.1016/0025-326X\(72\)90266-4](https://doi.org/10.1016/0025-326X(72)90266-4)
- 525 Elith, J., Graham, C.H., Anderson, R.P., Dudik, M., Ferrier, S., Guisan, A., Hijmans, R.J.,  
 526 Huettmann, F., Leathwick, J.R., Lehmann, A., Li, J., Lohmann, L.G., Loiselle, B.A.,  
 527 Manion, G., Moritz, C., Nakamura, M., Nakazawa, Y., Overton, J.M., Peterson, A.T.,  
 528 Phillips, S.J., Richardson, K., Scachetti-Pereira, R., Schapire, R.E., Soberon, J., Williams,  
 529 S., Wisz, M.S., Zimmermann, N.E., 2006. Novel methods improve prediction of species’  
 530 distributions from occurrence data. Ecography (Cop.). 29, 129–151.  
 531 <https://doi.org/10.1111/j.2006.0906-7590.04596.x>
- 532 Elith, J., Leathwick, J.R., Hastie, T., 2008. A working guide to boosted regression trees. J. Anim.  
 533 Ecol. 77, 802–813. <https://doi.org/10.1111/j.1365-2656.2008.01390.x>
- 534 Escobar, L.E., Qiao, H., Phelps, N.B.D., Wagner, C.K., Larkin, D.J., 2016. Realized niche shift  
 535 associated with the Eurasian charophyte *Nitellopsis obtusa* becoming invasive in North  
 536 America. Sci. Rep. 6, 1–15. <https://doi.org/10.1038/srep29037>
- 537 Falvey, M., Garreaud, R.D., 2009. Regional cooling in a warming world: Recent temperature  
 538 trends in the southeast Pacific and along the west coast of subtropical South America (1979-  
 539 2006). J. Geophys. Res. Atmos. 114, 1–16. <https://doi.org/10.1029/2008JD010519>

- Fielding, A.H., Bell, J.F., 1997. A review of methods for the assessment of prediction errors in conservation presence/absence models 24, 38–49.
- Figueroa, F.L., Santos, R., Conde-Álvarez, R., Mata, L., Gómez Pinchetti, J.L., Matos, J., Huovinen, P., Schuenhoff, A., Silva, J., 2006. The use of chlorophyll fluorescence for monitoring photosynthetic condition of two tank-cultivated red macroalgae using fishpond effluents. *Bot. Mar.* 49, 275–282. <https://doi.org/10.1515/BOT.2006.035>
- Freitas, C., Araújo, R., Bertocci, I., 2016. Patterns of benthic assemblages invaded and non-invaded by *Grateloupia turuturu* across rocky intertidal habitats. *J. Sea Res.* 115, 26–32. <https://doi.org/10.1016/j.seares.2016.07.002>
- Gattuso, J.-P., Gentili, B., Duarte, C.M., Kleypas, J.A., Middelburg, J.J., Antoine, D., 2006. Light availability in the coastal ocean: impact on the distribution of benthic photosynthetic organisms and contribution to primary production. *Biogeosciences Discuss.* 3, 895–959. <https://doi.org/10.5194/bgd-3-895-2006>
- Gouvêa, L.P., Schubert, N., Martins, C.D.L., Sissini, M., Ramlov, F., Rodrigues, E.R. de O., Bastos, E.O., Freire, V.C., Maraschin, M., Carlos Simonassi, J., Varela, D.A., Franco, D., Cassano, V., Fonseca, A.L., Barufi J, B., Horta, P.A., 2017. Interactive effects of marine heatwaves and eutrophication on the ecophysiology of a widespread and ecologically important macroalga. *Limnol. Oceanogr.* 62, 2056–2075. <https://doi.org/10.1002/lno.10551>
- Harlin, M.M., Villalard-Bohnsack, M., 2001. Seasonal dynamics and recruitment strategies of the invasive seaweed *Grateloupia doryphora* (Halymeniaceae, Rhodophyta) in Narragansett Bay and Rhode Island Sound, Rhode Island, USA. *Phycologia* 40, 468–474.

- 561 Hobday, A.J., Alexander, L. V., Perkins, S.E., Smale, D.A., Straub, S.C., Oliver, E.C.J.,  
 562 Benthuyssen, J.A., Burrows, M.T., Donat, M.G., Feng, M., Holbrook, N.J., Moore, P.J.,  
 563 Scannell, H.A., Sen Gupta, A., Wernberg, T., 2016. A hierarchical approach to defining  
 564 marine heatwaves. *Prog. Oceanogr.* 141, 227–238.  
 565 <https://doi.org/10.1016/j.pocean.2015.12.014>
- 566 Hoek, C. Van Den, 1982. The distribution of benthic marine algae in relation to the temperature  
 567 regulation of their life histories. *Biol J Linn Soc* 18, 81–144. [https://doi.org/10.1111/j.1095-](https://doi.org/10.1111/j.1095-8312.1982.tb02035.x)  
 568 [8312.1982.tb02035.x](https://doi.org/10.1111/j.1095-8312.1982.tb02035.x)
- 569 Hulme, P.E., 2009. Trade, transport and trouble: Managing invasive species pathways in an era  
 570 of globalization. *J. Appl. Ecol.* 46, 10–18. [https://doi.org/10.1111/j.1365-](https://doi.org/10.1111/j.1365-2664.2008.01600.x)  
 571 [2664.2008.01600.x](https://doi.org/10.1111/j.1365-2664.2008.01600.x)
- 572 Hulme, P.E., Bacher, S., Kenis, M., Klotz, S., Kühn, I., Minchin, D., Nentwig, W., Olenin, S.,  
 573 Panov, V., Pergl, J., Pyšek, P., Roques, A., Sol, D., Solarz, W., Vilà, M., 2008. Grasping at  
 574 the routes of biological invasions: A framework for integrating pathways into policy. *J.*  
 575 *Appl. Ecol.* 45, 403–414. <https://doi.org/10.1111/j.1365-2664.2007.01442.x>
- 576 Janiak, D.S., Whitlatch, R.B., 2012. Epifaunal and algal assemblages associated with the native  
 577 *Chondrus crispus* (Stackhouse) and the non-native *Grateloupia turuturu* (Yamada) in  
 578 eastern Long Island Sound. *J. Exp. Mar. Bio. Ecol.* 413, 38–44.  
 579 <https://doi.org/10.1016/j.jembe.2011.11.016>
- 580 Jones, E., Thornber, C.S., 2010. Effects of habitat-modifying invasive macroalgae on epiphytic  
 581 algal communities. *Mar. Ecol. Prog. Ser.* 400, 87–100. <https://doi.org/10.3354/meps08391>

- 582 Jueterbock, A., Smolina, I., Coyer, J.A., Hoarau, G., 2016. The fate of the Arctic seaweed *Fucus*  
 583 *distichus* under climate change: An ecological niche modeling approach. *Ecol. Evol.* 6,  
 584 1712–1724. <https://doi.org/10.1002/ece3.2001>
- 585 Jueterbock, A., Tyberghein, L., Verbruggen, H., Coyer, J.A., Olsen, J.L., Hoarau, G., 2013.  
 586 Climate change impact on seaweed meadow distribution in the North Atlantic rocky  
 587 intertidal. *Ecol. Evol.* 3, 1356–1373. <https://doi.org/10.1002/ece3.541>
- 588 Katsanevakis, S., Wallentinus, I., Zenetos, A., Leppäkoski, E., Çinar, M.E., Oztürk, B.,  
 589 Grabowski, M., Golani, D., Cardoso, A.C., 2014. Impacts of Invasive Alien Marine Species  
 590 on Ecosystem Services and Biodiversity: a pan-European Review. *Aquat. Invasions* 9, 391–  
 591 423. <https://doi.org/10.3391/ai.2014.9.4.01>
- 592 Kraemer, G., Yarish, C., Kim, J.K., Zhang, H., Lin, S., 2017. Life history interactions between  
 593 the red algae *Chondrus crispus* (Gigartinales) and *Grateloupia turuturu* (Halymeniales) in a  
 594 changing global environment. *Phycologia* 56, 176–185. <https://doi.org/10.2216/16-72.1>
- 595 Kramer-Schadt, S., Niedballa, J., Pilgrim, J.D., Schröder, B., Lindenborn, J., Reinfelder, V.,  
 596 Stillfried, M., Heckmann, I., Scharf, A.K., Augeri, D.M., Cheyne, S.M., Hearn, A.J., Ross,  
 597 J., Macdonald, D.W., Mathai, J., Eaton, J., Marshall, A.J., Semiadi, G., Rustam, R.,  
 598 Bernard, H., Alfred, R., Samejima, H., Duckworth, J.W., Breitenmoser-Wuersten, C.,  
 599 Belant, J.L., Hofer, H., Wilting, A., 2013. The importance of correcting for sampling bias in  
 600 MaxEnt species distribution models. *Divers. Distrib.* 19, 1366–1379.  
 601 <https://doi.org/10.1111/ddi.12096>
- 602 Lignell, A., Pedersén, M., 1989. Agar composition as a function of morphology and growth rate.  
 603 Studies on some morphological strains of *Gracilariaria secundata* and *Gracilariaria verrucosa*

- 604 (Rhodophyta). Bot. Mar. 32, 219–227. <https://doi.org/10.1515/botm.1989.32.3.219>
- 605 Liu, F., Pang, S.J., 2010. Stress tolerance and antioxidant enzymatic activities in the metabolisms  
606 of the reactive oxygen species in two intertidal red algae *Grateloupia turuturu* and  
607 *Palmaria palmata*. J. Exp. Mar. Bio. Ecol. 382, 82–87.  
608 <https://doi.org/10.1016/j.jembe.2009.11.005>
- 609 Marcelino, V.R., Verbruggen, H., 2015. Ecological niche models of invasive seaweeds. J.  
610 Phycol. 51, 606–620. <https://doi.org/10.1111/jpy.12322>
- 611 Martínez, B., Arenas, F., Trilla, A., Viejo, R.M., Carreño, F., 2015. Combining physiological  
612 threshold knowledge to species distribution models is key to improving forecasts of the  
613 future niche for macroalgae. Glob. Chang. Biol. 21, 1422–1433.  
614 <https://doi.org/10.1111/gcb.12655>
- 615 Martins, C.D.L., Arantes, N., Faveri, C., Batista, M.B., Oliveira, E.C., Pagliosa, P.R., Fonseca,  
616 A.L., Nunes, J.M.C., Chow, F., Pereira, S.B., Horta, P.A., 2012. The impact of coastal  
617 urbanization on the structure of phytobenthic communities in southern Brazil. Mar. Pollut.  
618 Bull. 64, 772–778. <https://doi.org/10.1016/j.marpolbul.2012.01.031>
- 619 Mathieson, A.C., Dawes, C.J., Pederson, J., Gladych, R.A., Carlton, J.T., 2008. The Asian red  
620 seaweed *Grateloupia turuturu* (Rhodophyta) invades the Gulf of Maine. Biol. Invasions 10,  
621 985–988. <https://doi.org/10.1007/s10530-007-9176-z>
- 622 Mulas, M., Bertocci, I., 2016. Devil's tongue weed (*Grateloupia turuturu* Yamada) in northern  
623 Portugal: Passenger or driver of change in native biodiversity? Mar. Environ. Res. 118, 1–9.  
624 <https://doi.org/10.1016/j.marenvres.2016.04.007>

- 625 Neiva, J., Assis, J., Coelho, N.C., Fernandes, F., Pearson, G.A., Serrão, E.A., 2015. Genes left  
626 behind: Climate change threatens cryptic genetic diversity in the canopy-forming seaweed  
627 *Bifurcaria bifurcata*. PLoS One 10, 1–21. <https://doi.org/10.1371/journal.pone.0131530>
- 628 Neiva, J., Assis, J., Fernandes, F., Pearson, G.A., Serrão, E.A., 2014. Species distribution models  
629 and mitochondrial DNA phylogeography suggest an extensive biogeographical shift in the  
630 high-intertidal seaweed *Pelvetia canaliculata*. J. Biogeogr. 41, 1137–1148.  
631 <https://doi.org/10.1111/jbi.12278>
- 632 Nyberg, C.D., Wallentinus, I., 2005. Can species traits be used to predict marine macroalgal  
633 introductions? Biol. Invasions 7, 265–279. <https://doi.org/10.1007/s10530-004-0738-z>
- 634 Occhipinti-Ambrogi, A., 2007. Global change and marine communities: Alien species and  
635 climate change. Mar. Pollut. Bull. 55, 342–352.  
636 <https://doi.org/10.1016/j.marpolbul.2006.11.014>
- 637 Pereira, M.D., Schettini, C.A.F., Omachi, C.Y., 2009. Caracterização de feições oceanográficas  
638 na plataforma de santa catarina através de imagens orbitais. Rev. Bras. Geofis. 27, 81–93.  
639 <https://doi.org/10.1590/S0102-261X2009000100007>
- 640 Peterson, T., 2003. Predicting the Geography of Species' Invasions via Ecological Niche  
641 Modeling. Q. Rev. Biol. 78, 419–433. <https://doi.org/10.1017/CBO9781107415324.004>
- 642 Phillips, S.J., Anderson, R.P., Schapire, R.E., 2006. Maximum entropy modeling of species  
643 geographic distributions. Ecol. Modell. 190, 231–259.  
644 <https://doi.org/10.1016/j.ecolmodel.2005.03.026>
- 645 Phillips, S.J., Dudík, M., Elith, J., Graham, C.H., Lehmann, A., Leathwick, J., Ferrier, S., 2009.

- 646 Sample selection bias and presence-only distribution models: Implications for background  
647 and pseudo-absence data. *Ecol. Appl.* 19, 181–197. <https://doi.org/10.1890/07-2153.1>
- 648 Poloczanska, E.S., Brown, C.J., Sydeman, W.J., Kiessling, W., Schoeman, D.S., Moore, P.J.,  
649 Brander, K., Bruno, J.F., Buckley, L.B., Burrows, M.T., Duarte, C.M., Halpern, B.S.,  
650 Holding, J., Kappel, C. V, O'Connor, M.I., Pandolfi, J.M., Parmesan, C., Schwing, F.,  
651 Thompson, S.A., Richardson, A.J., 2013. Global imprint of climate change on marine life.  
652 *Nat. Clim. Chang.* 3, 919–925. <https://doi.org/10.1038/Nclimate1958>
- 653 Saunders, G.W., Withall, R.D., 2006. Collections of the invasive species *Grateloupia turuturu*  
654 (Halymeniales, Rhodophyta) from Tasmania, Australia. *Phycologia* 45, 711–714.  
655 <https://doi.org/10.2216/06-10.1>
- 656 Schubert, N., García-Mendoza, E., Enríquez, S., 2011. Is the photo-acclimatory response of  
657 Rhodophyta conditioned by the species carotenoid profile? *Limnol. Oceanogr.* 56, 2347–  
658 2361. <https://doi.org/10.4319/lo.2011.56.6.2347>
- 659 Senay, S.D., Worner, S.P., Ikeda, T., 2013. Novel Three-Step Pseudo-Absence Selection  
660 Technique for Improved Species Distribution Modelling. *PLoS One* 8.  
661 <https://doi.org/10.1371/journal.pone.0071218>
- 662 Shea, K., 2002. Community ecology theory as a framework for biological invasions. *Trends*  
663 *Ecol. Evol.* 17, 170–176. [https://doi.org/10.1016/S0169-5347\(02\)02495-3](https://doi.org/10.1016/S0169-5347(02)02495-3)
- 664 Shea, K., Chesson, P., 2002. Community ecology theory as a framework for biological invasions.  
665 *Trends Ecol. Evol.* 17, 170–176. [https://doi.org/10.1016/s0169-5347\(02\)02495-3](https://doi.org/10.1016/s0169-5347(02)02495-3)
- 666 Simon, C., Ar Gall, E., Deslades, E., 2001. Expansion of the red alga *Grateloupia doryphora*

- 667 along the coasts of Brittany (France). *Hydrobiologia* 443, 23–29.  
 668 <https://doi.org/10.1023/A:1017587918604>
- 669 Sissini, M.N., de Barros Barreto, M.B.B., Széchy, M.T.M., de Lucena, M.B., Oliveira, M.C.,  
 670 Gower, J., Liu, G., de Oliveira Bastos, E., Milstein, D., Gusmão, F., Martinelli-Filho, J.E.,  
 671 Alves-Lima, C., Colepicolo, P., Ameka, G., de Graft-Johnson, K., Gouvea, L., Torrano-  
 672 Silva, B., Nauer, F., Marcos de Castro Nunes, J., Barufi, J.B., Rörig, L., Riosmena-  
 673 Rodríguez, R., Mello, T.J., Lotufo, L.V.C., Horta, P.A., 2017. The floating *Sargassum*  
 674 (Phaeophyceae) of the South Atlantic Ocean – likely scenarios. *Phycologia* 56, 321–328.  
 675 <https://doi.org/10.2216/16-92.1>
- 676 Sissini, M.N., Longo, G.O., Martins, C.D.L., Floeter, S.R., Pereira, S.B., Horta, P.A., 2014. First  
 677 record of the green alga *Halimeda* (Bryopsidales: Chlorophyta) at Rocas Atoll - Natural  
 678 dispersion or anthropogenic causes? *Mar. Biodivers. Rec.* 7.  
 679 <https://doi.org/10.1017/S1755267214001018>
- 680 Tsiamis, K., Aydogan, Ö., Bailly, N., Balistreri, P., Bariche, M., 2015. New Mediterranean  
 681 Biodiversity Records (July 2015). *Medit. Mar. Sci* 16, 472–488.  
 682 <https://doi.org/10.12681/mms.1440>
- 683 Tyberghein, L., Verbruggen, H., Pauly, K., Troupin, C., Mineur, F., De Clerck, O., 2012. Bio-  
 684 ORACLE: A global environmental dataset for marine species distribution modelling. *Glob.*  
 685 *Ecol. Biogeogr.* 21, 272–281. <https://doi.org/10.1111/j.1466-8238.2011.00656.x>
- 686 Ummenhofer, C.C., Meehl, G.A., 2017. Extreme weather and climate events with ecological  
 687 relevance: a review. *Philos. Trans. R. Soc. B Biol. Sci.* 372, 20160135.  
 688 <https://doi.org/10.1098/rstb.2016.0135>



- 689 Verbruggen, H., Tyberghein, L., Belton, G.S., Mineur, F., Jueterbock, A., Hoarau, G., Gurgel,  
690 C.F.D., De Clerck, O., 2013. Improving Transferability of Introduced Species' Distribution  
691 Models: New Tools to Forecast the Spread of a Highly Invasive Seaweed. PLoS One 8, 1–  
692 13. <https://doi.org/10.1371/journal.pone.0068337>
- 693 Verbruggen, H., Tyberghein, L., Pauly, K., Vlaeminck, C., Nieuwenhuyze, K. Van, Kooistra,  
694 W.H.C.F., Leliaert, F., de Clerck, O., 2009. Macroecology meets macroevolution:  
695 Evolutionary niche dynamics in the seaweed *Halimeda*. Glob. Ecol. Biogeogr. 18, 393–405.  
696 <https://doi.org/10.1111/j.1466-8238.2009.00463.x>
- 697 Vergés, A., Steinberg, P.D., Hay, M.E., Poore, A.G.B., Campbell, A.H., Ballesteros, E., Heck,  
698 K.L., Booth, D.J., Coleman, M.A., Feary, D.A., Figueira, W., Langlois, T., Marzinelli,  
699 E.M., Mizerek, T., Mumby, P.J., Nakamura, Y., Roughan, M., van Sebille, E., Gupta, A.  
700 Sen, Smale, D.A., Tomas, F., Wernberg, T., Wilson, S.K., 2014. The tropicalization of  
701 temperate marine ecosystems: climate-mediated changes in herbivory and community phase  
702 shifts. Proc. R. Soc. B Biol. Sci. 281, 1–10. <https://doi.org/10.1098/rspb.2014.0846>
- 703 Vilà, M., Basnou, C., Pyšek, P., Josefsson, M., Genovesi, P., Gollasch, S., Nentwig, W., Olenin,  
704 S., Roques, A., Roy, D., Hulme, P.E., 2010. How well do we understand the impacts of  
705 alien species on ecosystem services? A pan-European, cross-taxa assessment. Front. Ecol.  
706 Environ. 8, 135–144. <https://doi.org/10.1890/080083>
- 707 Wenger, S.J., Olden, J.D., 2012. Assessing transferability of ecological models: An  
708 underappreciated aspect of statistical validation. Methods Ecol. Evol. 3, 260–267.  
709 <https://doi.org/10.1111/j.2041-210X.2011.00170.x>
- 710 Wiens, J.A., Stralberg, D., Jongsomjit, D., Howell, C.A., Snyder, M.A., 2009. Niches, models,

and climate change: Assessing the assumptions and uncertainties. *Proc. Natl. Acad. Sci.* 106, 19729–19736. <https://doi.org/10.1073/pnas.0901639106>

Williams, S.L., Smith, J.E., 2007. A Global Review of the Distribution, Taxonomy, and Impacts of Introduced Seaweeds. *Annu. Rev. Ecol. Evol. Syst.* 38, 327–359. <https://doi.org/10.1146/annurev.ecolsys.38.091206.095543>

Figure captions

**Fig. 1** Map of *G. turuturu* occurrences used in the model. Sampling locations are represented in white diamonds, whilst black dots represent occurrences in the known introduced range and grey dots represent occurrences in the native range. A close-up was made in South America to show more clearly the introduced and sampled locations in this region.

**Fig. 2** Map of *G. turuturu* habitat suitability in the planet (a), with zoom in South America (b), and Europe/North Africa (c). Regions in black represent regions suitable for the occurrence of *G. turuturu*, whilst regions in light grey represent regions with unsuitable environmental conditions for *G. turuturu*. Land is represented in dark grey, and zones without environmental data are represented in white (no data).

**Fig. 3** Response curve of *G. turuturu* to maximum ocean temperature. Decline point is set at 21.23°C and the limiting point at 26.55°C, demonstrating low habitat suitability of *G. turuturu* to higher temperatures.

**Fig. 4** Average  $\pm$  standard deviation ( $n=7$ ) of Optimal Quantum Yield (Fv/Fm) for all temperatures (18, 24 and 30°C) and nutrient conditions (low nutrient condition – LNC, and high nutrient concentration – HNC) in the first, third, fifth and seventh day of the warm temperate adapted population (WAP) experiment. Letters indicate Tukey's test significantly different groups ( $p<0.05$ ).

**Fig. 5** Average  $\pm$  standard deviation ( $n=6$ ) of Optimal Quantum Yield (Fv/Fm) in the temperate adapted population (TAP) experiment in the beginning of the experiment (initial control), then the values of each treatment after three days at 10 (control), 13, 16, 20 and 24°C. Letters indicate Tukey's test significantly different groups ( $p<0.05$ ).

**Fig. 6** Response of *G. turuturu* (warm temperate adapted population - WAP) to different temperatures in treatments. Different letters represent Tukey's test significantly ( $p<0.05$ ) different groups. Bars represent average  $\pm$  standard deviation. (a) Chlorophyll *a* concentration ( $\mu\text{g}$  chlorophyll-*a* per g of fresh weight) in the experiment's beginning and end ( $n=7$ ). (b) Total Carotenoids concentration ( $\mu\text{g}$  total carotenoids per g of fresh weight) in experiment's beginning and end ( $n=7$ ). (c) Percentage of *G. turuturu* daily growth rate during the experiment ( $n=10$ ).

**Fig. 7** Average  $\pm$  standard deviation ( $n=7$ ) of phycoerythrin concentration ( $\mu\text{g}$  phycoerythrin per g of fresh weight) at different nutrient conditions (warm temperate adapted population - WAP). Letters indicate Tukey's test significantly different groups ( $p<0.05$ ).

Supplementary material

**ESM1:** Excel file with all the occurrences and their sources used in the Ecological Niche Model.

752   **ESM2:** Word document with additional figures, such as the experimental design setup, response  
753   curves of the model and chlorophyll concentration in each nutrient condition (WAP experiment).

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